

**SPECIFICATION AMENDMENTS**

**Please replace paragraphs number [0012]-[0013] on page 4, with the following**

**rewritten paragraphs:**

[0012] FIG. 1 is a serum sodium calibration curve. The calibration curve was generated using the methods disclosed in the Example 1. Briefly, the calibration curve was constructed by ~~plotting~~ plotting the  $\Delta A$  values of the standards against the corresponding sodium concentration.

[0013] FIG. 2 is a serum lithium calibration curve. The calibration curve was generated using the methods disclosed in the Example 2. Briefly, the calibration curve was constructed by ~~plotting~~ plotting the  $\Delta A$  values of the standards against the corresponding lithium concentration.

**Please replace paragraph number [00112] on pages 26-27, with the following rewritten**

**paragraph:**

[00112] **Assay Principle.** Sodium was determined spectrophotometrically through a kinetic coupling assay system involving the chimeric 3'(2'),5'-bisphosphate nucleotidase (as described in Section B) whose activity was sensitive to sodium concentration ( $IC_{50}=20mM$ ). Through enzymatic coupling, the phosphatase substrate, adenosine 3',5'-bisphosphate (PAP) was converted to hypoxanthine by a series of enzymatic reactions to generate uric acid and hydrogen peroxide ( $H_2O_2$ ).  $H_2O_2$  generated reacts with N-Ethyl-N-(2-hydroxy-3-sulfopropyl)-3-m-toluidine (EHSPT) and 4-aminoantipyrine (4-AA) in the presence of peroxidase (POD) to form a quinone dye which had maximal absorbance at 556nm. The rate of the quinone dye formation was inversely proportion to the concentration of ~~lithium~~ sodium in serum samples. The enzymatic coupling reaction scheme is shown below in Table 3:

**Please replace Table 6 on page 30, with the following rewritten Table:**

**TABLE 6**

Phosphatase	
PAP + H <sub>2</sub> O	-----→ AMP + Pi
5'-Nucleotidase/ADA deaminase/PNP	
AMP	-----→ Hypoxanthine + Pi + NH <sub>3</sub> + R-1-P
Xanthine Oxidase	
Hypoxanthine + 2H <sub>2</sub> O + 2O <sub>2</sub>	-----→ Uric Acid + 2H <sub>2</sub> O <sub>2</sub>
Peroxidase	
H <sub>2</sub> O <sub>2</sub> + 4-AA + EHSPT	-----→ 4H <sub>2</sub> O + Quinone dye (λ max 556nm)

PAP: 3'-phosphoadenosine 5'-phosphate (adenosine 3',5'-bisphosphate)

AMP: Adenosine-5'-phosphate

PNP: Purine Nucleoside Phosphorylase

4-AA: 4-Aminoantipyrine

EHSPT: N-Ethyl-N-(2-Hydroxy-3-Sulfopropyl)-m-Toluidine ~~Toluidine~~

**Please replace paragraph [0040] on pages 10-11, with the following rewritten**

**paragraph:**

[0040] Any suitable 3',5' bisphosphate nucleotidase can be used. In one example, the 3',5' bisphosphate nucleotidase is of *Saccharomyces cerevisiae* origin (See e.g., Murguía et al., *J. Biol. Chem.*, 271(46):29029-33 (1996)). This nucleotidase is also known as the HAL2 nucleotidase. Moreover, any suitable 3',5' bisphosphate nucleotidase catalyzing the reaction defined in Section B can be used in the present compositions and methods. The enzyme useful in the present compositions and methods is not limited those enzymes having only 3'(2'),5'-bisphosphate nucleotidase activity. For example, the enzyme may have dual enzymatic activity, e.g., Tol-1. Homologues of the HAL2 phosphatase are also contemplated. Useful enzymes capable of catalyzing the above reaction include, but are not limited to BPntase (see, e.g., Spiegelberg et al., *J. Biol. Chem.* 274(19):13619-28 (1999)), HsPIP, RnPIP (see, e.g., López-Coronado, et al., *J. Biol. Chem.* 274(23):16034-39 (1999), and Tol-1 (see, e.g., ~~Amoto~~ Miyamoto, et al., *J. Bacteriol.* 182(13):3619-25 (2000)). Other useful 3'(2'),5'-bisphosphate

nucleotidases, *e.g.*, 3',5' bisphosphate nucleotidases are disclosed in Peng et al., *J. Biol. Chem.* 270(49):29105-10 (1995), Dichtl et al., *EMBO J.*, 16(23):7184-95 (1997), Gil-Mascarell (1997), Gil-Mascarell et al., *The Plant J.* 17(4):373-83 (1999), ~~can also be used~~. A functional fragment or a derivative of a 3'(2'),5'-bisphosphate nucleotidase that still substantially retain its enzymatic activity catalyzing the dephosphorylation of adenosine 3',5'-bisphosphate to yield corresponding adenosine 5'-phosphate (AMP) and P<sub>i</sub> can also be used.

**Please replace paragraph [0069] on page 18, with the following rewritten paragraph:**

[0069] H<sub>2</sub>O<sub>2</sub> formation can be assessed ~~any~~ assessed by any suitable means. In one embodiment, the H<sub>2</sub>O<sub>2</sub> formation is assessed by a peroxidase and Trinder reaction. Any suitable peroxidase can be used. More preferably, a horseradish peroxidase is used. For example, the horseradish peroxidases with the following GenBank accession Nos. can be used: E01651; D90116 (prxC3 gene); D90115 (prxC2 gene); J05552 (Synthetic isoenzyme C(HRP-C)); S14268 (neutral); OPRHC (C1 precursor); S00627 (C1C precursor); JH0150 (C3 precursor); S00626 (C1B precursor); JH0149 (C2 precursor); CAA00083 (*Armoracia rusticana*); and AAA72223 (synthetic horseradish peroxidase isoenzyme C (HRP-C)). Any suitable Trinder reagent can be used herein. Hydrogen peroxide can be quantitated by the quinone dye assay. *See, e.g., Tamaøkel Tamaoku, et al., Chem. Pharm. Bull.* 30: 2497 (1982); Shimojo et al., *Clin. Chem.* 35(9):1992-94 (1989). The amount of quinone dye formed is inversely related to the amount of sodium ions in the sample.